New establishment of annexes for Testing Methods for Fertilizers

Upon revising to the "Testing Methods for Fertilizers (2021)", following suit the example set by the Japanese Industrial Standards (JIS), content that is difficult to be included in descriptions of a testing method due to, for example, a large amount of information is to be newly arranged as annexes.

When an annex provides standards, it is indicated as (Standards). When an annex provides reference information, it is indicated as (Reference).

• Annexes to be newly established

Annex A (Standards) Procedure to validate characteristics of testing methods

In the Testing Methods for Fertilizers (2020), this was provided as an appendix. In this version, this is to be provided as Annex A.

Annex B (Reference) List of extraction methods for main components, etc.

For the same extraction procedures for main components, etc., general descriptions from water-soluble to total are provided in the comments for the testing method that is explained first (e.g., phosphoric acid) in the Testing Methods for Fertilizers, and the readers are referred to the general descriptions in the comments for subsequent testing methods (e.g., potassium). In the Testing Methods for Fertilizers (2021), the relevant comments are to be revised to refer to Annex B instead.

Annex C1 (Reference) and Annex C2 (Reference) Standard solution, etc. preparation methods in simultaneous analysis using ICP optical emission spectrometer and ICP mass spectrometer

For the preparation methods of mixed standard solutions in simultaneous analysis using ICP optical emission spectrometer and ICP mass spectrometer, all necessary information such as preparation concentrations of standard solutions is provided in the comments for the testing method that is explained first (e.g., phosphoric acid) in the Testing Methods for Fertilizers, and the readers are referred to the relevant descriptions in the comments for subsequent testing methods (e.g., potassium). In the Testing Methods for Fertilizers (2021), the relevant comments are to be revised to refer to Annex C1 or C2 instead.

Annex D (Reference) Example IC column used for acid-soluble sulfur

Even if the functional group is the same, the bonding state of the functional group, degree of cross-linking, etc. varies depending on the model of the column. In order to separate the ion to be measured, measurement conditions suitable for the column to be used (e.g., composition of mobile phase (eluent), gradient) are applied. For that reason, a list of measurement conditions which were validated through collaborative studies, etc. are shown in Annex D.

Annex A (Standards)

The procedure to validate characteristics of testing methods

(1) **Purposes**

This article explains the procedure to validate characteristics of testing methods which will be listed in the Testing Methods of Fertilizers. In addition, when testing institutes conduct a test which is not included in the Testing Methods of Fertilizers, a procedure to evaluate the validity of the test method should conform to a method stipulated in this article.

Additionally, this article targets chemical testing methods. However, this article is not applicable to the extraction method of the content of effective figures (acid-, citric acid- and water-soluble) in a powdery sample or a solidified fertilizer.

Comment 1 The contents of effective figures (acid-, citric acid- and water-soluble) are stipulated in a notification of the Ministry of Agriculture, Forestry and Fisheries. In addition, the change of measurement conditions such as an extraction temperature may affect an observed value in some cases. Therefore, no changes will be implemented in the extraction method of the contents of effective figures in a powdery fertilizer and a solidified fertilizer for the present and the application of this article is limited to the change of a measurement method (including refining of extract, etc.).

(2) **Definition of terminology** 1, 2, 3, 4, 5, 6, 7

The definition of terminology in this article is as shown below.

- a) Selectivity: Capability to accurately measure components subjected to analysis under the existence of materials which seem to exist in a sample.
- **b**) **Trueness**: The degree of agreement between the mean obtained from multiple measurement results and the true value ⁽¹⁾.
- c) **Precision**: The degree of agreement (or the degree of variation) among the independent measurement results which are repeatedly measured under the determined conditions.
- d) **Repeatability**: The precision of the measurement results of analytical samples, which are regarded to be all identical, obtained under condition (repeatability conditions) that independent measurement results are measured in a short time, using the same method, in the same laboratory, by the same operator and with the same instrument.
- e) Intermediate precision: The precision of a measurement result of analytical samples, which are regarded to be all identical, obtained under condition (intermediate conditions) that independent measurement results are measured, using the same method, in the same laboratory and in different factors (such as different time and a different operator).
- f) Reproducibility: The precision of a measurement result of analytical samples, which are regarded to be all identical, obtained under condition (reproducibility conditions) that independent measurement results are measured, using the same method, in different laboratories, by different operators and with different instruments.
- g) Minimum Limit of Quantification (LOQ): The quantifiable lowest volume or minimum concentration of a component subjected to analysis which is contained in an analytical sample.
- h) Minimum Limit of Detection (LOD): The detectable lowest volume or minimum concentration of a component subjected to analysis which is contained in an analytical sample.
- i) **Reference material**: A material which is uniform and stable enough for one or more prescribed properties, and is made suitable for the purpose of use in a measurement process.
- **j**) **Certified reference material**: A reference material, whose values of one or more prescribed properties are characterized by a reasonable metrological procedure, having a certificate of attestation on which the characteristics of prescribed properties and their uncertainty and metrological traceability are stated.
- **k**) **Blank sample**: An analytical sample not containing components subjected to analysis ⁽²⁾.
- **I)** Addition sample: An analytical sample the content of whose components subjected to analysis is known, or an analytical sample to which reference materials are added ^{(3) (4)} or compounded ⁽³⁾.
- m) Natural contamination sample: A test sample prepared from fertilizers which naturally contain the components subjected to analysis such as harmful components.
- **n**) **Distribution sample**: An analytical sample prepared from fertilizers ⁽⁵⁾ which are manufactured in a fertilizer production factory, etc.

- o) Surrogate: A material which is added to an analytical sample in order to conduct a pre-process operation, correct yields in respective steps of measurement procedure and confirm recovery rates, whose chemical structure is identical or similar to a target component.
- **p**) **SN ratio**: Intensity ratio of a signal (response value) S originating from the analysis target and a signal (usually noise) N based on the other factors.
 - **Note** (1) In reality, the certified value of a certified reference material, the chemical composition of a compound, the added content of a reference material, etc. and others.
 - (2) Reagents, etc. containing a target matrix can be used in the case that there is no distribution fertilizer used as a blank sample for a recovery test and the confirmation of the minimum limit of quantification, etc.
 - (3) Mix a component subjected to analysis with a mortar, etc. to sufficient uniformity
 - (4) In the case of adding a standard solution, vaporize the solvent sufficiently conducting measures such as letting it stand for one night.
 - (5) A fertilizer containing components subjected to analysis whose formation or form changed due to a chemical or physical process (a granulation process, etc.).

(3) Validation method

Test necessary items of (3.1) to (3.8) in a planned manner and estimate performance parameters from the obtained results.

Confirm whether the estimated values of performance parameters are suitable to target values (performance norm) respectively, and evaluate that the test method is validated if they are all suitable.

(3.1) Scope of application

As a result of a validation test in a single laboratory and a collaborative study, if the result is suitable up to reproducibility, the test method is evaluated as a validated test method as far as the kind of a fertilizer used in the test and the range of concentration are concerned. Therefore, a laboratory where the said test is conducted can use the performance (reproducibility, etc.) as a validated method through implementing internal quality control, etc.

As a result of a validation test in a single laboratory, if the result is suitable to trueness, repeatability and intermediate precision, etc., the test method is evaluated as a validated test method as far as the laboratory where the test was conducted and as far as the kind of a fertilizer used in the test and the range of concentration are concerned. Therefore, another laboratory which wants to introduce the test method is required to carry out the validation anew in an individual laboratory with the above test method.

(3.2) Selectivity ^{8, 9, 10, 11})

(3.2.1)Case of Chromatography

Conduct a procedure for a blank sample and confirm that there is no peak (interference peak) which affects the measurement of components subjected to analysis ⁽⁶⁾. In addition, in the case of the simultaneous measurement of multi components, confirm that adjacent peaks are sufficiently separated ⁽⁶⁾.

Note (6) Resolution (*R*) should be 1.0 or more at minimum though 1.5 or more is preferable.

Comment 2 Resolution (R) is used as a separation indicator of peaks. If Resolution (R) is 1.5 or more, the adjacent two peaks are sufficiently separated and they do not affect a measurement, whether a peak height or peak area is used. If Resolution (R) is 1.0 or more, there is no problem if peak height is used for a measurement even if the adjacent two peaks overlap to some extent.

Resolution (R) can be obtained using a peak width by the formula (1a). In addition, if the peak is a normal distribution, it can be obtained using a peak width at half height by the formula (1b). With the data processing device of a chromatograph, the formula (1b) is often used to obtain Resolution (R).

Resolution (R) =
$$\frac{t_2 - t_1}{\frac{1}{2} \times (W_1 + W_2)} \qquad \cdots (1a)$$
$$1.18 \times (t_2 - t_1)$$

Resolution (R) =
$$\frac{1.18 \wedge (l_2 - l_1)}{\left(\frac{W_1}{2^{-1}} + \frac{W_1}{2^{-2}}\right)}$$
(1b)

t_I : Retention time of Peak 1	t_2 : Retention time of Peak 2
W_1 : Peak width of Peak 1	W_2 : Peak width of Peak 1

 $W_{\frac{1}{2},1}$: Peak width at half height of Peak 1

 $W_{\underline{1},2}$: Peak width at half height of Peak 2

(3.2.2)Case of a method other than Chromatography⁽⁷⁾

Conduct a procedure for a blank sample and confirm that there is no response which originates from other components than a component subjected to analysis and can be a factor of positive error of a quantification value ⁽⁸⁾.

Note (7) A test method such as Molecular absorption spectrometry, Atomic absorption spectrometry or Titration analysis which does not isolate with a measurement instrument.

(8) Absorbance, titer, etc.

(3.3) Calibration curve ^{8, 12, 13)}

Measure respective standard solutions for the calibration curve preparation of the concentration or the content ⁽⁹⁾ of level 6 to 8 a few times ⁽¹⁰⁾ to make a figure plotting the obtained signals ⁽¹¹⁾ as a function of the concentration or the content of a component subjected to analysis and evaluate its linearity visually using the figure.

If linearity is recognized, calculate the inclination (*b*) and the intercept (*a*) of a calibration curve, its confidence interval and the coefficient of determination (r^2) using a statistical method such as the calculation of a regression equation by the least square method. Moreover make the plot of residuals ⁽¹²⁾ in respective levels.

- Note (9) The blank test solution for the calibration curve preparation can be included.
 - (10) In order to avoid nonlinear confusion due to the variation of sensitivity, etc., conduct measurements randomly for each replicate determination.
 - (11) Absorbance, fluorescence intensity, peak height, peak area, etc.
 - (12) The difference between a signal obtained by measurement and a signal estimated using a regression equation.

Comment 3 It is recommended that the 95% confidence interval of an intercept (*a*) includes the origin (0).

Comment 4 Though it is usable if the coefficient of determination (r^2) is 0.99 or more, it is recommended that the coefficient of determination (r^2) is 0.999 or more for a precise analysis. If it is less than 0.99, use the equation of a higher order or consider changing the measuring area.

Comment 5 The mean of residuals is 0 and the residuals indicate a random pattern.

(3.4) Trueness ^{7, 8, 12, 14, 15)}

As the estimation method of trueness, the methods are recommended in the following order. (1) Use of a certified

reference material (3.4.1), (2) Comparison with an observed value by a validated method (3.4.2) and (3) Recovery test (3.4.3).

In addition, if a surrogate is used, it is recommended that a recovery rate is about 40% or more.

(3.4.1)Use of a certified reference material

With regard to a component which has matrixes similar to a fertilizer subjected to test and can use a certified reference material containing components subjected to measurement of the concentration in a measurement level, conduct repeatability testing using 3 or more analytical samples (n) according to the test method of the certified reference material. As a result, the mean of the observed values should be within the warning limit to the certified value (characteristic value) or the absolute value of the difference between the mean of the observed values and the certified value (characteristic value) should not exceed 2 times of the standard uncertainty composed of respective uncertainties of the mean of the observed values and the certified value ($^{(13)}$.

Comment 6 A warning limit is given using the formula (2) which is obtained from a collaborative study for the characterization of a certified reference material.

A warning limit for a certified value (μ)

$$=\mu\pm 2\times \sqrt{(s_R^2-s_r^2)+\frac{s_r^2}{n}}=\mu\pm 2\times \sqrt{s_L^2+\frac{s_r^2}{n}}\qquad \cdots (2)$$

- μ : Certified value
- s_R : Reproducibility standard deviation in a collaborative study
- s_r : Repeatability standard deviation in a collaborative study ⁽¹⁴⁾
- *n*: The number of analytical samples to repeatability test
- s_L : Pure between-laboratory standard deviation in a collaborative study
- Note (13) The evaluation procedure of the difference between a measurement result and a certified value (characteristic value) is shown in Reference 1 Procedure to compare an observed value and a certified value.
 - (14) It may be expressed as within-laboratory standard deviation (s_W) in some cases.

(3.4.2)Use of another validated test method

For a component for which a certified reference material is not usable but another validated test method (hereinafter referred to as "a standard test method") is applicable, confirm that the condition \mathbf{a}) or \mathbf{b}) is satisfied.

a) In case 12 or more samples are available: Conduct respective tests of 12 or more test samples composed of addition samples, natural contamination samples or distribution samples according to a new test method and a standard test method, create the correlation chart of observed values with two methods for each sample and calculate the inclination (*b*) and the intercept (*a*) of a regression line, and a correlation coefficient (*r*). Further confirm a prediction interval.

However, in case that the width between the minimum and the maximum observed value is small, conduct the paired samples *t*-test to confirm that a significant difference is not observed.

- **Comment 7** It is recommended that the 95% confidence interval of an inclination (*b*) includes 1, the 95% confidence interval of an intercept (*a*) includes the origin (0) and the correlation coefficient (*r*) is no less than 0.99.
- b) In case fewer samples are available: With regard to 3 or more test samples of different concentration, conduct respective repeatability addition tests using 4 analytical samples according to the new test method and a standard test method, confirm the homoscedasticity of the results of 2 groups and conduct a *t*-test for each concentration to confirm that significant difference is not observed under the two-sided significant level

of 5%.

(3.4.3) In case neither certified reference material nor other validated test methods are usable

For 3 or more test samples of different concentration, conduct respective repeatability tests using 3 analytical samples and evaluate by obtaining the recovery using the mean of the observed values. The criteria of the trueness are shown in **Separate sheet: The target of trueness and the criteria of precision in respective concentration levels**.

(**3.5**) **Precision** ^{8, 12, 16, 17})

Evaluate reproducibility and repeatability by a collaborative study (3.5.1). Or evaluate an intermediate precision and repeatability by a repeatability test (3.5.2).

(3.5.1)Reproducibility and repeatability by a collaborative study

The number of laboratories to obtain effective data should be 8 or more ⁽¹⁵⁾. Conduct undisclosed duplicate collaborative studies for 5 or more kinds of samples with different concentration. Obtain reproducibility and repeatability from the observed values ⁽¹⁶⁾ to evaluate.

Criteria to evaluate these precisions are shown in Separate sheet: The target of trueness and the criteria of precision in respective concentration levels.

- Note (15) In case the number of laboratories which have required facility/instruments is limited, this should be 5 or more.
 - (16) The calculation method is shown in **Reference 2: Calculation of reproducibility or intermediate** precision and repeatability

(3.5.2)Intermediate precision and repeatability by a repeatability test in a single laboratory on different days

Conduct a duplicate test ⁽¹⁷⁾ per test day for 5 to 7 days using two analytical samples of different concentration which is included in a normal range ⁽¹⁸⁾. Obtain intermediate precision and repeatability from the observed values ⁽¹⁹⁾ to evaluate.

Criteria to evaluate these precisions are shown in Separate sheet: The target of trueness and the criteria of precision in respective concentration levels.

- Note (17) The data of internal quality control can be used.
 - (18) It is not necessary for the same tester to conduct a test through 5 to 7 days.
 - (19) The calculation method is shown in **Reference 2: Calculation of reproducibility or intermediate** precision and repeatability

(3.6) Minimum Limit of Quantification (LOQ) ^{7, 11)}

Estimate Minimum Limit of Quantification according to (3.6.1) to (3.6.3). Prepare test samples which include the concentration estimated to be near the Minimum Limit of Quantification step by step as necessary. And conduct respective repeatability tests using 3 analytical samples and define the concentration of a prepared test sample as the Minimum Limit of Quantification, where the mean of the obtained values using the prepared test sample is suitable to the target value of trueness.

- **Comment 8** In case permissible content and equivalent level is 1.0 mg/kg or more, the Minimum Limit of Quantification (LOQ) of harmful components and restricted components, etc. should be no more than 1/5 of the permissible content and equivalent level. In case permissible content and equivalent level is no more than 1.0 mg/kg, the Minimum Limit of Quantification (LOQ) should be no more than 2/5 of the permissible content. Moreover, it is recommended that the Minimum Limit of Quantification of main components/major components and material components should be no more than 1/5 of minimum volume to be contained and the minimum content of a distribution fertilizer. In addition, in case the Minimum Limit of Quantification exceeds 1/5 of these minimum volumes, conduct the above-mentioned repeatability test, confirm the Minimum Limit of Quantification and state clearly the fact in the applicable range of a test method.
- Comment 9 There are some methods to estimate Minimum Limit of Quantification. The methods differ depending on whether they are based on an instrument analysis or not and depending on instruments used. A method different from the methods shown in (3.6.1) to (3.6.3) is allowed. However, the definition of a method

and Minimum Limit of Quantification by the method should be clearly stated.

(3.6.1)Estimation method by a repeatability test

With regard to a test sample with concentration near Minimum Limit of Quantification, conduct a repeatability test using 7 to 10 analytical samples, obtain repeatability standard deviation and estimate Minimum Limit of Quantification (LOQ) in an analytical sample by the formula (3).

Estimated value of Minimum Limit of Quantification (LOQ) = $10 \times s_r \qquad \cdots (3)$

 s_r : Repeatability standard deviation

(3.6.2) Estimation method using a calibration curve

In case a calibration curve is linear, estimate Minimum Limit of Quantification (LOQ) in an analytical sample by the formula (4) using the standard deviation of the residuals of a calibration curve or estimated signals in concentration 0 and the inclination of a calibration curve.

Estimated value of Minimum Limit of Quantification (LOQ) = $\frac{10 \times s}{b}$...(4)

- s: The standard deviation of residuals. Or the standard deviation of signasls in concentration 0, which are estimated from a regression line
- b: The inclination of a calibration curve

(3.6.3)Estimation method using an SN ratio

In a test method such as Chromatography, etc. which has a baseline noise, calculate from a concentration in an analytical solution whose SN ratio is 10 to 1 at the peak and estimate Minimum Limit of Quantification (LOQ) in an analytical sample.

(3.7) Minimum Limit of Detection (LOD) ^{7, 11)}

Estimate Minimum Limit of Detection according to (3.7.1) to (3.7.3).

Comment 10 There are some methods to estimate Minimum Limit of Detection. The methods differ depending on whether they are based on an instrument analysis or not and depending on instruments used. A method different from the methods shown in (3.7.1) to (3.7.3) is allowed. However, the definition of a method and Minimum Limit of Detection by the method should be clearly stated.

(3.7.1)Estimation method by a repeatability test

With regard to a test sample or a blank sample with concentration near Minimum Limit of Quantification, conduct repeatability tests using 7 to 10 analytical samples, obtain repeatability standard deviation and estimate Minimum Limit of Detection (LOD) in an analytical sample by the formula (5).

Estimated value of Minimum Limit of Detection (LOD) in an analytical sample = $2 \times t(n-1, 0.05) \times s_r \qquad \cdots (5)$

 s_r : Repeatability standard deviation

t (n-1,0.05): The Student value of Significance Level 5% (one side) ⁽²⁰⁾

n: The number of analytical samples in a repeatability test

Note (20) In case of a repeatability test using 7 analytical samples, the value is 1.94. In case of using 10

analytical samples, the value is 1.83.

(3.7.2)Estimation method using a calibration curve

In case a calibration curve is linear, estimate Minimum Limit of Detection (LOD) in an analytical sample by the formula (6) using the standard deviation of the residuals of a calibration curve or estimated signals in concentration 0 and the inclination (b) of a calibration curve.

Estimated value of Minimum Limit of Detection (LOD)

$$=\frac{2\times t(n-2,\,0.05)\times s}{b}\qquad \cdots (6)$$

- s: The standard deviation of residuals. Or the standard deviation of signals in concentration 0, which are estimated from a regression line
- b: The inclination of a calibration curve
- t(n-2, 0.05): The Student value of Significance Level 5% (one side)
- *n*: The number of a measurement point on a calibration curve

(3.7.3) Estimation method using an SN ratio

In a test method such as Chromatography, etc. which has a baseline noise, calculate from a concentration in an analytical solution whose SN ratio is 3 to 1 at the peak and estimate Minimum Limit of Detection (LOD) in an analytical sample.

(3.8) **Robustness** ^{7, 11, 12)}

Robustness should be studied when an analysis method is developed, and the estimation method depends on the type of analysis method to be developed. Robustness expresses the reliability of an analysis method when its analysis conditions are intentionally changed. If an observed value tends to be easily affected by the variation of an analysis condition, it is necessary to consider a method to control an analysis condition appropriately or to state the fact as a precaution in a testing method. The evaluation of robustness enables the establishment of a series of parameters such as Resolution related to system conformance. Similarly, the confirmation of these parameters ensures that the validation of an analysis method is maintained in a daily analysis.

Typical variation factors are as follows.

(3.8.1)Common variation factors: Typical variation factors common to various kinds of test methods are as follows.

- a) Extraction time, extraction temperature
- b) Stability of a test solution in respective steps
- c) Reagent's grade

(3.8.2) Variation factors in Chromatography, etc.: Typical variation factors of measurements by Chromatography or refining by solid phase extraction are as follows.

- a) Change of a column or a cartridge (A different lot or a different brand)
- b) Influence by the variation of pH and composition of an eluent or a wash
- c) Temperature
- d) Flow rate
- e) Influence of a matrix and effect of dilution

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Reference 1: Procedure to compare an observed value and a certified value

Obtain the total mean (m) of the replication test results and the certified value (μ), and the absolute value (Δ_m) of the difference of the two values by the formula (R1.1). Next, obtain the standard uncertainty (u_{CRM}) of the certified value of a certified reference material by the formula (R1.2), and obtain the standard uncertainty (u_m) of the total mean by the formula (R1.3). Calculate the combined standard uncertainty ($u_{C(\Delta_m)}$) of Δ_m by the formula (R1.4) using the obtained u_m and u_{CRM} . Further, calculate an expanded uncertainty (U_{Δ_m}) by the formula (R1.5) using the coverage factor (k = 2).

Compare Δ_m and U_{Δ_m} to confirm that the criterion (the formula (R1.6)) is satisfied, that is, Δ_m is no more than U_{Δ_m} .

The absolute value (Δ_m) of the difference of the total mean of repeatability test results and a certified value= $|m-\mu|$ ···(R1.1)

The standard uncertainty (u_{CRM}) of the certified value= $\frac{U_{95\%}}{k_{CRM}}$ ···(R1.2)

The standard uncertainty of the measurement of a total mean $(u_m) = \frac{s_r}{\sqrt{n}}$...(R1.3)

The combined standard uncertainty $(u_{C(\Delta_m)})$ of $\Delta_m = \sqrt{u_m^2 + u_{CRM}^2}$...(R1.4)

The expanded uncertainty (U_{Δ_m}) of $\Delta_m = k_{C(\Delta_m)} \times u_{C(\Delta_m)} = 2 \times u_{C(\Delta_m)}$ (R1.5)

Criterion $\Delta_m \leq U_{\Delta_m} \cdots (R1.6)$

m: The total mean of observed values

 μ : A certified value

 $U_{95\%}$: The expanded uncertainty of a certified value

 k_{CRM} : The coverage factor of an expanded uncertainty of a standard reference material

- s_r : Repeatability standard deviation
- *n*: The number of repeatability test samples
- $k_{C(\Delta_m)}$: The coverage factor of an expanded uncertainty of Δ_m ($k_{C(\Delta_m)}=2$)

Reference 2: Calculation of reproducibility or intermediate precision and repeatability

(1) Structure of an observed value

An observed value (x_{ij}) in Table 1, as is shown in the formula (R2.1), consists of a true value (μ) , a variation (β) by a factor and a variation (e) by an accidental error under repeatability conditions (hereinafter referred to as "an accidental error"). When *p* laboratories conduct a collaborative study in which respective laboratories conduct repeatability tests using *n* samples, the formula (R2.2) is introduced on the assumption that the distribution of β is equivalent to $N(0, \sigma_L^2)$ which depends on a pure between-laboratory variation and the distribution of e is equivalent to $N(0, \sigma_r^2)$ which depends on an accidental error. In addition, when the same laboratory conducts replicate tests for *p* days using *n* samples on respective test days, the formula (R2.3) is introduced on the assumption that the distribution of β is equivalent to $N(0, \sigma_{(T)}^2)$ which depends on test days variation (factor *T*) and the distribution of e is equivalent to $(0, \sigma_r^2)$ which depends on an accidental error.

Observed value $(x_{ij}) = \mu + \beta_i + e_{ij}$ ··· (R2.1) Observed value $(x_{ij}) = \mu + N(0,\sigma_L^2) + N(0,\sigma_r^2)$ ··· (R2.2) Observed value $(x_{ij}) = \mu + N(0,\sigma_{(T)}^2) + N(0,\sigma_r^2)$ ··· (R2.3)

 μ : True value

 β_i : Variation of a factor e_{ij} : Accidental error

 $N(0,\sigma_L^2)$: Normal distribution of β_i with the mean 0 and standard deviation σ_{L_i}

 $N(0,\sigma_r^2)$: Normal distribution of e_{ij} with the mean 0 and standard deviation σ_r

 σ_L^2 : Pure between - laboratory variance σ_r^2 : Repeatability variance

 $N(0, \sigma_{(T)}^2)$: Normal distribution of β_i with the mean 0 and standard deviation $\sigma_{(T)}$

 $\sigma_{(T)}^2$: Test days variance

Table 1 Test results of collaborative studies or repeatability tests on different days

Laboratory or test day	Analytical sample number						
(Factor)	1	2	3	•••	j	•••	n
1	x 11	x 12	x 13	•••	x 1j	•••	x_{1n}
2	x 21	X_{22}	X 23	•••	x 2j	•••	x_{2n}
3	x 31	X_{32}	x 33	•••	x 3j	•••	x_{3n}
•••	•••	•••	•••	•••	•••	•••	•••
i	x_{i1}	X_{i2}	x 13	•••	x _{ij}	•••	x in
•••	•••	•••	•••	•••	•••	•••	•••
p	x_{p1}	X_{p2}	x _{p3}	•••	x _{pj}	•••	x _{pn}

(2) Calculation procedure of reproducibility and repeatability of the results of a collaborative study

(2.1) Estimation of true value and variance

In an actual statistical analysis, a true value (μ), a true and pure between-laboratory variance (σ_L^2) and a true repeatability variance (σ_r^2) are unknown. Therefore, they are replaced with estimated values obtained from the results of a collaborative study and are expressed as a mean (m), a pure between-laboratory variance (s_L^2) and a repeatability variance (s_r^2) respectively.

(2.2) One-way analysis of variance

Exclude ineffective observed values which have clearly objective reasons such as deviation from a protocol and

malfunction of instruments from the report values by laboratories which participated in a collaborative study. Further exclude outliers by conducting Cochran's test and Grubb's test. And conduct one-way analysis of variance for the remaining results to obtain the unbiased variance (V) of respective variation factors in Table 2.

	Table 2 Table o	f one-way analy	sis of variance	
Variation factor	Sum of squares	Dgree of	Unbiased	Expectation of variance
	(S)	freedom (φ)	variance (V)	(E(V))
Between-laboratory (L)	SSL	p-1	V_{L}	$\sigma_r^2 + n \times \sigma_L^2$
Accidental error (e)	SSr	$p \times (n \neg 1)$	Vr	σ_r^2

Comment 1 It is possible to conduct one-way analysis of variance easily using a statistical program or a tool of a spreadsheet program. In this case, it should be noted that different terminologies may be used (Between-laboratory $(L) \rightarrow$ Between-group, Accidental error $(e) \rightarrow$ Within-group, Unbiased variance \rightarrow Mean square, etc.).

Comment 2 Unbiased variance (V) is calculated by (Sum of squares)/ (Degree of freedom).

(2.3) The calculation of reproducibility and repeatability

The relation of the expectation of variance E (*V*) of respective factors in Table 2 holds true. Therefore, calculate repeatability variance (s_r^2) and pure between-laboratory variance (s_L^2) by the formula (R2.4) and (R2.5), and further calculate reproducibility variance (s_R^2) by the formula (R2.6) ^{(1) (2)}.

Repeatability variance $(s_r^2) = V_r$ ··· (R2.4) Pure between - laboratory variance $(s_L^2) = \frac{V_L - V_r}{n}$ ··· (R2.5) Reproducibility variance $(s_R^2) = s_L^2 + s_r^2$ ··· (R2.6)

- V_r : The unbiased variance of a variation factor (accidental error (e)) in the table of one – way analysis of variance (Table 2)
- V_L : The unbiased variance of a variation factor (between laboratories (*L*)) in the table of one – way analysis of variance (Table 2)

Calculate a repeatability standard deviation (s_r) and a reproducibility standard deviation (s_R) by the formula (R2.7) and (R2.8) using the obtained repeatability variance and reproducibility variance, and further calculate a repeatability relative standard deviation (RSD_r) and a reproducibility relative standard deviation (RSD_R) by the formula (R2.9) and (R2.10) $^{(2)}$ (³).

Repeatability standard deviation $(s_r) = \sqrt{s_r^2}$ ··· (R2.7) Reproducibility standard deviation $(s_R) = \sqrt{s_R^2}$ ··· (R2.8) Repeatability relative standard deviation $(RSD_r, \%) = \frac{s_r}{m} \times 100$ ··· (R2.9) Reproducibility relative standard deviation $(RSD_R, \%) = \frac{s_R}{m} \times 100$ ··· (R2.10)

m: the gross mean of the effective data of collaborative study results

Note (1) In case $V_L < V_r$, assume $V_L = V_r$ (that is, the pure between-laboratory variance $(s_L^2) = 0$ in the formula (R2.5)) and let the formula (R2.6) form $s_R^2 = s_r^2$.

- (2) The rounding of a numerical value is not executed in the middle of the calculation.
- (3) The mean and the standard deviation are expressed rounding to the digit of the observed value. The relative standard deviation is expressed rounding to the first decimal place.

(3) Calculation procedure of intermediate precision and repeatability by the replicate test results on different days

(3.1) Estimation of a true value and a variance

In an actual statistical analysis, a true value (μ), a true test day variance ($\sigma_{(T)}^2$) and a true repeatability variance (σ_r^2) are unknown. Therefore, they are replaced with estimated values obtained from the repeatability test results on different days and are expressed as a mean (m), test days variance ($s_{(T)}^2$) and a repeatability variance (s_r^2) respectively.

(3.2) One-way analysis of variance

Conduct one-way analysis of variance for the replicate test results on different days to obtain the unbiased variance (V) of respective variation factors in Table 3.

	Table 3 Table o	f one-way analy	sis of variance	
Variation factor	Sum of squares	Dgree of	Unbiased	Expectation of variance
Variation factor	(S)	freedom (φ)	variance (V)	(E(V))
Test days (T)	SS _T	p-1	VT	$\sigma_r^2 + n \times \sigma_{(T)}^2$
Accidental error (e)	SSr	$p \times (n \neg 1)$	Vr	σ_r^2

Comment 3 It is possible to conduct one-way analysis of variance easily using a statistical program or a tool of a spreadsheet program. In this case, it should be noted that different terminologies may be used (Test days $(T) \rightarrow$ between-group, Accidental error $(e) \rightarrow$ within-group, Unbiased variance \rightarrow Mean square, etc.). **Comment 4** Unbiased variance (V) is calculated by (Sum of squares)/ (Degree of freedom).

(3.3) The calculation of intermediate precision and repeatability

The relation of the expectation of variance E (V) of respective factors in Table 3 holds true. Therefore, calculate repeatability variance (s_r^2) and test days variance $(s_{(T)}^2)$ by the formula (R2.11) and (R2.12), and further calculate intermediate variance $(s_{I(T)}^2)$ by the formula (R2.13) ^{(2) (4)}.

- Repeatability variance $(s_r^2) = V_r \cdots$ (R2.11) Test days variance $(s_{(T)}^2) = \frac{V_T - V_r}{n} \cdots$ (R2.12) Intermediate Variance $(s_{I(T)}^2) = s_{(T)}^2 + s_r^2 \cdots$ (R2.13)
 - V_r : The unbiased variance of a variation factor (accidental error (e))

in the table of one - way analysis of variance (Table 3)

 V_T : The unbiased variance of a variation factor (test days (T))

in the table of one – way analysis of variance (Table 3)

Calculate a repeatability standard deviation (s_r) and an intermediate standard deviation $(s_{I(T)})$ by the formula (R2.14) and (R2.15) using the obtained estimated values of repeatability variance and intermediate variance, and further calculate a repeatability relative standard deviation (RSD_r) and an intermediate relative standard deviation $(RSD_{I(T)})$ by the formula (R2.16) and (R2.17) $^{(2)}(3)$.

Repeatability standard deviation $(s_r) = \sqrt{s_r^2}$ ··· (R2.14) Intermediate standard deviation $(s_{I_{(T)}}) = \sqrt{s_{I_{(T)}}^2}$ ··· (R2.15)

Repeatability relative standard deviation $(RSD_r, \%) = \frac{s_r}{m_{\times 100}}$ (R2.16) Intermediate relative standard deviation $(RSD_{I_{(7)}}, \%) = \frac{s_I}{m_{\times 100}}$ (R2.17)

m: Gross mean of the replicate test results on different days

Note (4) In case $V_T < V_r$ assume $V_T = V_r$ (that is, the test days variance $(s_{(T)}^2)$ in the formula (R2.12) = 0) and let the formula (R2.13) form $s_{I(T)}^2 = s_r^2$.

(4) Examples of the calculation of intermediate precision and repeatability by the replicate test results on different days.

An example of repeatability test results on different days of citric acid-soluble phosphoric acid using sample 1 and sample 2 containing phosphite is shown in Table 4. Conduct one-way analysis of variance for the test results of respective samples to obtain the unbiased variance (V) of respective variation factors (Table 5).

Examples of the calculation of intermediate precision and repeatability for the sample 1 and sample 2 using the formula (R2.11) to the formula (R2.17) are shown in Table 6-1 and 6-2. In addition, the results of respective standard deviation are expressed rounding to the digit of the observed value and the results of the respective relative standard deviations are expressed rounding to the first decimal place.

	Table 4 Example of repeatability test results on different days					(mass fraction (%))		
	Test day (factor)							Total mean
Sample No	1	2	3	4	5	6	7	$(m)^{1)}$
Sample 1	51.20	52.15	51.00	51.35	51.35	51.38	51.28	51.38
Sample 1	51.45	51.85	51.09	51.28	51.10	51.38	51.43	51.56
Sample 2	5.18	4.90	5.01	5.15	5.14	5.13	5.21	5 10
Sample 2	5.00	5.12	5.06	5.14	5.07	5.11	5.18	5.10

1) The mean is expressed rounding to the digit of the observed value.

		Table 5 Table of one-way analysis of variance			
Sample No.	Variation factor	Sum of squares	Dgree of	Unbiased	Expectation of variance
Sample No	Variation factor	<i>(S)</i>	freedom (φ)	variance (V)	(E(V))
Sample 1	Test days (T)	1.0570	6	0.17616	$\sigma_r^2 + n \times \sigma_{I(T)}^2$
	Accidental error (e)	0.1253	7	0.01789	σ_r^2
Sample 2	Test days (T)	0.0478	6	0.00797	$\sigma_r^2 + n \times \sigma_{I(T)}^2$
	Accidental error (e)	0.0448	7	0.00640	σ_r^2

Variation factor	Formula	Calculation	Result
Repeatability variance (s_r^2)	= V _r	= 0.01789	0.01789
Repeatability standard deviation $(s_r)^{2}$	$=\sqrt{{s_r}^2}$	$=\sqrt{0.01789}$	0.13 (%) ⁴⁾
Repeatability relative standard deviation $(RSD_r)^{3)}$	$= (s_r/m) \times 100$	= (0.1338/51.38)×100	0.3 (%)
Teat days variance $(s_{(T)}^2)$	$= (V_{\rm T} \neg V_{\rm f})/n$	= (0.17616-0.01789)/2	0.07914
Intermediate variance $(s_{I(T)}^2)$	$= s_{T}^{2} + s_{r}^{2}$	= 0.07914+0.01789	0.09703
Intermediate standard deviation $(s_{I(T)})^{2}$	$=\sqrt{{s_{I(T)}}^2}$	$=\sqrt{0.09703}$	0.31 (%) ⁴⁾
Intermediate relative standard deviation $(RSD_{ICD})^{3}$	$= (s_{I(T)}/m) \times 100$	= (0.3115/51.38)×100	0.6 (%)

Table 6-1 Calculation of intermediate precision and repeatability using the sample 1 replicate test results on different days¹⁾

1) The rounding of a numerical value is not executed in the middle of the calculation.

2) The standard deviation is expressed rounding to the digit of the observed value.

3) The relative standard deviation is expressed rounding to the first decimal place.

4) Mass fraction

Table 6-2 Calculation of intermediat	e precision a	nd repeatability	using the	sample 2	replicate	test resul	ts on
different davs1)							

Variation factor	Formula	Calculation	Result
Repeatability variance (s_f^2)	$= V_{\rm f}$	= 0.00640	0.00640
Repeatability standard deviation $(s_r)^{2}$	$=\sqrt{s_r^2}$	$=\sqrt{0.00640}$	0.08 (%) ⁴⁾
Repeatability relative standard deviation $(RSD_r)^{3)}$	$= (s_r/m) \times 100$	= (0.0800/5.10)×100	1.6 (%)
Teat days variance $(s_{(T)}^2)$	$= (V_{\rm T} \neg V_{\rm f})/n$	= (0.00797-0.00640)/2	0.00078
Intermediate variance $(s_{I(T)}^2)$	$= s_{T}^{2} + s_{r}^{2}$	= 0.00078+0.00640	0.00718
Intermediate standard deviation $(s_{I(T)})^{2}$	$=\sqrt{s_{I(T)}^2}$	$=\sqrt{0.00718}$	0.08 (%) ⁴⁾
Intermediate relative standard deviation $(RSD_{I(T)})^{3}$	$= (s_{I(T)}/m) \times 100$	= (0.0848/5.10)×100	1.7 (%)
Footpote: Pofer to Table 6 1			

Footnote: Refer to Table 6-1

Separate sheet: The target of trueness and the criteria of precision in respective concentration levels

The target of trueness (recovery rate) and the criteria of precision in respective concentration levels to evaluate Chromatography ⁽¹⁾ and test methods other than Chromatography are shown in Table 1 and Table 2. The target of trueness is generally within the recovery rate of Table 1. As for precision, the permissible level may exceed respective relative standard deviations in Table 2 by a factor of 2.0.

Note (1) Gas chromatography, Gas Chromatography/Mass Spectrometry, High-Performance Liquid Chromatography, High-Performance Liquid Chromatography/Tandem Mass Spectrometry, Ion Chromatography, etc.

Table 1 The target of trueness in respective concentration levels					
	Chromatography	Test methods other than			
	515	Chromatography			
Concentration level	Recovery rate (%)	Recovery rate (%)			
≥25 % (mass fraction)	90~108	98~102			
≥ 10 % (mass fraction)	90~108	97~103			
≥1 % (mass fraction)	85~110	96~104			
≥ 0.1 % (mass fraction)	85~110	94~106			
≧100 mg/kg	80~115	92~108			
≧10 mg/kg	70~120	90~110			
≧1 mg/kg	70~120	85~115			
≧100 μg/kg	70~120	85~115			
≧10 μg/kg	70~120	80~120			
<10 µg/kg	60~125	75~125			

Table 2 Criteria of p	precision ¹⁾ in	respective	concentration i	levels
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	Chromatography			Test methods other than Chromatography			
Concentration level	Reproducibility relative standard deviation	Intermediate relative standard deviation	Repeatability relative standard deviation	Reproducibility relative standard deviation	Intermediate relative standard deviation	Repeatability relative standard deviation	
	(%)	(%)	(%)	(%)	(%)	(%)	
\geq 25 % (mass fraction)	8	6.5	4	2.5	2	1	
\geq 10 % (mass fraction)	8	6.5	4	3	2.5	1.5	
\geq 1 % (mass fraction)	8	6.5	4	4	3.5	2	
≥ 0.1 % (mass fraction)	8	6.5	4	6	4.5	3	
≧100 mg/kg	8	6.5	4	8	6.5	4	
≧10 mg/kg	11	9	6	11	9	6	
≧1 mg/kg	16	13	8	16	13	8	
≧100 µg/kg	22	18	11	22	18	11	
≧10 μg/kg	22	18	11	22	18	11	
<10 µg/kg	22	18	11	22	18	11	

1) As for precision, the permissible level may exceed respective relative standard deviations by a factor of 2.0.

Annex B

(Reference)

List of extraction methods for main components, etc.

(1) List of extraction methods for main components, etc.

Lists of extraction methods for main components, etc. used in the Testing Methods for Fertilizers are shown in Table 1 to Table 4.

			Extraction method ^{a)}							
Component name		Test method	Kjeldahl digestion	Incineration- hydrochloric acid boiling	Incineration- aqua regia digestion	Nitric acid, sulfuric acid, perchloric acid digestion	Microwave headed acid digestion	Other		
Total	4.1.1.a	Kjaeldahl method	○ ^{b)}							
nitrogen	4.1.1.b	Combustion method						А		
	4.1.1.c	Devarda's alloy – Kjeldahl method						В		
	4.1.1.d	Reduced iron - Kjeldahl method						С		
Total phosphoric acid	4.2.1.a	Ammonium vanadomolybdate absorptiometric analysis	0	0	0					
	4.2.1.b	Quinoline gravimetric analysis	0							
Total	4.3.1.a	Flame atomic absorption								
1 otal		spectrometry or flame		0	0					
porassium		photometry								
	4.3.1.b	Sodium tetraphenylborate								
		gravimetric analysis		0						
Total lime	451a	Flame atomic absorption		0	\bigcirc					
i otur inne	1.5.1.4	spectrometry		0	0					
Total	4.6.1.a	Flame atomic absorption		\cap	\bigcirc					
magnesia	nonna	spectrometry		0	0					
Total zinc	4.9.1.a	Flame atomic absorption		\cap	\bigcirc					
		spectrometry		0	0					
	4.9.1.b	ICP Optical Emission			\cap					
		Spectrometry			0					
Total copper	4.10.1.a	Flame atomic absorption		0	\cap					
		spectrometry								
	4.10.1.b	ICP Optical Emission			\cap			1		
	4.10.1.0	Spectrometry			\cup					

Table 1 List of extraction methods for main components, harmful components (e.g., heavy metals), etc.

a) Respective extraction method

A: Introduction of analytical sample into measuring instrument

B: Reduction by Devarda's alloy - Kjeldahl digestion

C: Reduction by reduced iron – Kjeldahl digestion

D: Dichromate oxidation

E: Hydrochloric acid treatment

F: Extraction with water and sulfuric acid (1+5)

G: Boiling with potassium hydroxide/ethanol solution

H: Nitric acid, perchloric acid digestion

I: Decomposition at microwave decomposition temperature of 240 $^{\circ}\mathrm{C}$

J: Nitric acid, sulfuric acid, perchloric acid digestion with added ammonium sulfate

K: Ammonium hydrogensulfate melting

b) Filtration is not necessary

		I		Extraction mathed ^{a)}						
					Extractio	n method"				
Component name				Incineration	Incineration	Nitric acid,	Microwave			
		Test mathed	Kjeldah1	hudeashtasis	and an	sulfuric acid,	handad said	Other		
		1 est method	digestion	nydroenione	aqua regia	perchloric	neaded acto	Other		
			_	acid boiling	digestion	acid digestion	digestion			
Organic	4.11.1.a	Dichromate oxidation						D		
carbon	41115	Combustion method						F		
Total milfur	4.11.1.0	Potacium cormanganata						2		
I OTAL SULLY	4.12.1.a	Potassium permanganate						F		
content		analysis								
	4.12.1.b	Barium chloride gravimetric						G		
		analysis								
	4.12.1.e	Transmitted light analysis						G		
Mercury	512	Cold vapor atomic absorption						н		
		spectrometry								
		Cold vapor atomic absorption								
	5.1.b	spectrometry (Fluid sludge					I			
		fertilizers)								
		Hydride generation atomic				-				
Arsenic	5.2.a	absorption spectrometry				0				
		Silver diethyl dithiocarbamate								
	5.2.b	shoestionatsia analysis				0				
	5.2 -									
	0.2.C	ICP Mass Spectrometry					0			
Cadmium	5.3.a	Flame atomic absorption			0					
		spectrometry								
	535	ICP Optical Emission			0					
	2.2.0	Spectrometry			Ŭ					
	5.3.c	ICP Mass Spectrometry					0			
Mintert	5.4.a	Flame atomic absorption								
Nickel		spectrometry			0					
		ICP Optical Emission			_			•••••		
	5.4.b	Spectrometry			0					
	540	ICP Mass Spectrometry					0			
	2.4.0	Flame atomic absorption								
Character	55-	Fiame atomic absorption								
Chromium	0.0.a	spectrometry (Fertilizers			l v					
		containing organic matters)								
		Flame atomic absorption								
	5.5.b	spectrometry (Fertilizers						J		
		mainly containing fused								
		matters, slag, etc.)								
		Flame atomic absorption								
	5.5.c	spectrometry (Fertilizers not				0				
		containing organic matters)								
		ICP Optical Emission			~					
	5.5.6	Spectrometry								
	••••••	ICP Mass Spectrometry								
	550	(Fartilizars containing organic					0			
	0.0.0	(rettinzers containing organic					Ū			
		Flama atomic abagenting								
Lead	5.6.a	Flame atomic absorption			0					
		spectrometry								
	5.6.b	ICP Optical Emission			0					
		Spectrometry								
	5.6.c	ICP Mass Spectrometry					0			
Titanium	511-	ICP Optical Emission				0				
- Italiloili	2.11.4	Spectrometry (1)				Ĭ				
	5116	ICP Optical Emission						V		
	5.11.6	Spectrometry (2)						K		
		Flame atomic absorption		-						
Sodium	8.4.a	spectrometry		0						
				1						

Table 1 (cont.)

				Extraction	n method ^{a)}	
Component name		Test method	Boiling	Constant- temperature vertical rotating mixer	Horizontal reciprocating water bath shaker	Other
Citrate- soluble phosphoric acid	4.2.2.a	Ammonium vanadomolybdate absorptiometric analysis				А
	4.2.2.b	Quinoline gravimetric analysis				А
Acid and	4.4.1.a	Potassium fluoride method		0	0	
base-soluble silicic acid	4.4.1.b	Potassium fluoride method (Silica gel fertilizers, etc.)				В
	4.4.1.c	Potassium fluoride method (Fertilizers containing silica gel fertilizers)				С
	4.4.1.d	Perchloric acid method		0		
Acid-soluble lime	4.5.2.a	Flame atomic absorption spectrometry	0			
Alkalinity	4.5.5.a	Ethylenediamine tetraacetate method	0			
Acid-soluble magnesia	4.6.2.a	Flame atomic absorption spectrometry	0			
Acid-soluble manganese	4.7.1.a	Flame atomic absorption spectrometry	0			
Acid-soluble sulfur	4.12.2.a	Ion Chromatography				D

 Table 2
 List of extraction methods for soluble main components, etc.

a) Respective extraction method

A: Heating in Petermans citrate solution after separating water-soluble components with water

B: Heating in sodium hydroxide solution (20 g/L)

C: Heating in sodium hydroxide solution (20 g/L) after separating acid-soluble components by heating in hydrochloric acid (1+23)

D: Extraction with hydrochloric acid (1+23)

		Extraction method			
Component name		Test method	Constant- temperature vertical rotating mixer	Horizontal reciprocating water bath shaker	
Citric acid- soluble phosphoric acid	4.2.3.a	Ammonium vanadomolybdate absorptiometric analysis	0	0	
	4.2.3.b	Ammonium vanadomolybdate absorptiometric analysis (Fertilizers containing phosphorous acid or phosphite)	0	0	
	4.2.3.c	Quinoline gravimetric analysis	0		
	4.2.3.d	ICP Optical Emission Spectrometry	0	0	
Citric acid- soluble potassium	4.3.2.a	Flame atomic absorption spectrometry or flame photometry	0	0	
	4.3.2.b	Sodium tetraphenylborate gravimetric analysis	0		
	4.3.2.c	Sodium tetraphenylborate volumetric analysis	0		
	4.3.2.d	ICP Optical Emission Spectrometry	0	0	
Citric acid-	4.5.3.a	Flame atomic absorption spectrometry	0	0	
soluble lime	4.5.3.b	ICP Optical Emission Spectrometry	0	0	
Citric acid- soluble	4.6.3.a	Flame atomic absorption spectrometry	0	0	
magnesia	4.6.3.b	ICP Optical Emission Spectrometry	0	0	
Citric acid- soluble	4.7.2.a	Flame atomic absorption spectrometry	0	0	
manganese	4.7.2.b	ICP Optical Emission Spectrometry	0	0	
Citric acid- soluble boron	4.8.1.a 4.8.1.b	Azomethine-H method ICP Optical Emission Spectrometry	0	0	

Table 3 List of extraction methods for citric acid-soluble main components

			Extraction method ^{a)}						
Component name		Test method	Vertical rotating mixer	Vertical reciprocating shaker	Boiling	Shaking to mix ^{b)}	Other		
Ammoniacal	4.1.2.a	4.1.2.a Distillation method		A			В		
nitrogen	4.1.2.b	Formaldehyde method	С						
Nitrate	4.1.3.a	Devarda's alloy - distillation method					В		
nitrogen	4.1.3.b	Reduced iron- distillation method					В		
-	4.1.3.c	Phenol sulfuric acid method	D	D		D			
Water-soluble phosphoric acid	4.2.4.a	Ammonium vanadomolybdate absorptiometric analysis	0	0		0			
4.2.4.b	4.2.4.b	Ammonium vanadomolybdate absorptiometric analysis (Fertilizers containing phosphorous acid or phosphite)	0	0		0			
	4.2.4.c	Quinoline gravimetric analysis	0						
	4.2.4.d	ICP Optical Emission Spectrometry	0	0		0			
Water-soluble potassium	4.3.3.a	Flame atomic absorption spectrometry or flame photometry	0	0	0	0			
	4.3.2.b	Sodium tetraphenylborate gravimetric analysis	0		0				
	4.3.2.c	Sodium tetraphenylborate volumetric analysis	0		0				
	4.3.2.d	ICP Optical Emission Spectrometry	0	0	0	0			
Water-soluble silicic acid	4.4.2.a	Potassium fluoride method	0			Е			
Water-soluble	4.5.4.a	Flame atomic absorption spectrometry	F			0			
calcium	4.5.3.b	ICP Optical Emission Spectrometry				0			
Water-soluble	4.6.4.a	Flame atomic absorption spectrometry			G	0			
magnesia	4.6.4.b	ICP Optical Emission Spectrometry			G	0			
Water-soluble	4.7.3.a	Flame atomic absorption spectrometry	0	0		0			
manganese	4.7.3.b	ICP Optical Emission Spectrometry	0	0		0			
Watan 1-1-1	4.8.2.a	Azomethine-H method		1	0	0			
boron	4.8.2.b	ICP Optical Emission Spectrometry			0	0			

Table 4 List of extraction methods for water-soluble main components, etc.

a) Respective extraction method

A: Extraction with hydrochloric acid (1+23)

B: Introduction of analytical sample into distillation apparatus

C: Extraction with hydrochloric acid (1+20)

D: Extraction with copper sulfate - silver sulfate solution

E: 5 g of analytical sample - 500 mL of water

F: 1 g of analytical sample - 500 mL of water

G: 1 g of analytical sample - 500 mL of water

b) Extraction method for liquid fertilizers, 1 g of analytical sample - 100 mL of water

				Extra	ction method	a)	
Component name		Test method	Vertical rotating mixer	Vertical reciprocating shaker	Boiling	Shaking to mix ^{b)}	Other
Water-soluble	4.9.2.a	Flame atomic absorption spectrometry	0			0	
zine	4.9.2.b ICP Optical Emission Spectrometry					0	
Water-soluble copper	4.10.1.a	Flame atomic absorption spectrometry	0			0	
	4.10.2.Ъ	ICP Optical Emission Spectrometry				0	
Water-soluble iron	4.13.1.a	Flame atomic absorption spectrometry	0			0	
	4.13.1.b	ICP Optical Emission Spectrometry				0	
Water-soluble molybdenum	4.14.1.a	Sodium thiocyanate absorptiometric analysis	0			0	
	4.14.1.b	ICP Optical Emission Spectrometry				0	
Water-soluble	4.15.1.a	Flame atomic absorption spectrometry				0	
cobalt	4.15.1.b	ICP Optical Emission Spectrometry				0	

T.1.1.1/

Annex C1

(Reference)

List of concentration ranges of calibration curve in ICP optical emission spectrometry

(1) List for ICP optical emission spectrometry

A list of concentration ranges of calibration curve in ICP optical emission spectrometry used in the Testing Methods for Fertilizers is shown in Table 1.

Note that when preparing mixed standard solutions, avoid using standard solutions made of compounds (potassium dihydrogenphosphate etc.) which contain anything other than a target component (phosphoric acid, etc.) or prepare separately the standard solution for calibration curve of a component (potassium, etc.) which affects measurement. When preserving mixed standard solutions for calibration curve preparation, use a container, which can be sealed tightly, made of material such as PTFE that does not elute boron easily.

			Conce	ntration range			
Component name	Test method	Element concentration (mg/L)		Conversion factor	Oxide concentration (mg/L		Measurement wavelength (nm)
Phosphoric acid	4.2.3.d, 4.2.4.d	Р	1~200	2.2914	P_2O_5	2.291~458.2	178.287
Potassium	4.3.2.d, 4.3.3.d	Κ	1~200	1.2046	K ₂ O	1.205~241.0	766.491, 769.896
Lime (calcium)	4.5.3.b, 4.5.4.b	Ca	0.1~20	1.3992	CaO	0.1399~27.98	393.366, 317.933
Magnesia	4.6.3.b, 4.6.4.b	Mg	0.1~20	1.6583	MgO	0.1658~33.16	279.553, 280.270
Manganese	4.7.2.b, 4.7.3.b	Mn	0.05~10	1.2912	MnO	0.06455~12.91	257.610, 260.569
Boron	4.8.1.b, 4.8.2.b	В	0.05~10	3.2199	B_2O_3	0.1610~32.20	249.773, 249.678
Zinc	4.9.2.b	Zn	0.1~20	—		—	213.856, 206.200
Copper	4.10.2.b	Cu	0.1~20	—		—	327.396, 224.700, 324.754
Iron	4.13.2.b	Fe	0.1~20	—		—	259.940, 238.204
Molybdenum	4.14.1.b	Mo	0.1~20	_		_	202.030, 277.540
Cobalt	4.15.1.b	Co	0.1~20	_		_	228.616

Table 1 Concentration range of calibration curve and measurement wavelength for each element in ICP Optical Emission Spectrometry

Annex C2

(Reference)

List of concentration ranges of calibration curve in ICP mass spectrometry

(1) List for ICP mass spectrometry

A list of concentration ranges of calibration curve in ICP mass spectrometry used in the Testing Methods for Fertilizers is shown in Table 1.

		IVI	ass spectron	letry			
Component name		Co	oncentration r calibration c	ange of urve	Internal standard element		
	Test method	Element and concentration (µg/L)		Mass number	Elen conc	nent and entration ug/L)	Mass number
Arsenic	5.2.c	As	0.2~20	75	Te	10	125
Cadmium	5.3.c	Cd	0.05~5	111	Rh	5	103
Nickel	5.4.c	Ni	0.5~50	60	Rh	5	103
Chromium	5.5.e	Cr	1~100	52	Sc	50	45
Lead	5.6.c	Pb	0.1~10	208	T1	5	205

 Table 1 Concentration range of calibration curve and mass number for each element in ICP

 Mass Spectrometry

Annex D

(Reference)

Example IC column used for acid-soluble sulfur (4.12.2.a Ion Chromatography)

This Annex shows the ion exchange column and measurement conditions used for acid-soluble sulfur (4.12.2.a Ion Chromatography), as summarized in Table 1.

Example IC chromatograms of standard solution, compound fertilizer sample solution and gypsum sample solution produced by the measurement conditions provided in Table 1 are shown in Figure 1 to Figure 4. In the figures, the peak of sulfate iron (SO₄²⁻) is indicated by an arrow (\downarrow).

	Ion exchange column		Measurement conditions					
Functional group	IC column name	Inner diameter × Length Particle diameter	Eluent and eluting conditions	Flow rate	Sample injection volume	Column temperature	Note	
Quaternary ammonium group	Shodex IC SI-90 4E	4.0 mm×250 mm, 9 μm	1.8-mM sodium carbonate - 1.7-mM sodium hydrogencarbonate solution	1.0 mL/min	20 µL	25°C	А	
Quaternary ammonium group	Metrosep A Supp 4- 250/4.0	4.0 mm×250 mm, 9 μm	1.8-mM sodium carbonate - 1.7-mM sodium hydrogencarbonate solution	1.0 mL/min	20 µL	Room temperature	_	
						25°C	В	
Quaternary ammonium group	Shim-pack IC-SA2	4.0 mm×250 mm, 9 μm	1.8-mM sodium carbonate - 1.7-mM sodium hydrogencarbonate solution	1.0 mL/min	20 µL	25°C	С	
Quaternary ammonium group	PCI-205	4.0 mm×250 mm, 9 μm	1.8-mM sodium carbonate - 1.7-mM sodium hydrogencarbonate solution	1.0 mL/min	20 µL	37°C	_	
Quaternary ammonium	TSKgel SuperIC-Anion	4.6 mm×100 mm,	0.8-mM sodium carbonate - 7.5-mM sodium hydrogencarbonate solution	1.5	30 µL	40%0	_	
group	HS	3.5 µm	1.1-mM sodium carbonate - 7.5-mM sodium hydrogencarbonate solution	1.5 mL/min		40°C	_	
Quaternary ammonium group	TSKgel SuperIC-AZ	4.6 mm×150 mm, 4 μm	1.7-mM sodium carbonate - 6.3-mM sodium hydrogencarbonate solution	0.8 mL/min	20 µL	40°C	D	
Quaternary alky lamines	IonPac AS12A	4.0 mm×200 mm, 9 μm	2.7-mM sodium carbonate - 0.3-mM sodium hydrogencarbonate solution	1.2 mL/min	20 µL	32°C	_	
		4.0 mm×250 mm.		1.0 mL/min	50 µL		_	
Quaternary alkanolamines	IonPac AS22	6 µm	4.5-mM sodium carbonate - 1.4-mM sodium hydrogencarbonate solution	1.2 mL/min	25 μL	- 35°C	_	
Quaternary alkanolamines	IonPac AS19	4.0 mm ×250 mm, 7.5 μm	10-mM potassium hydroxide solution	1.0 mL/min	20 µL	30°C	_	
Quaternary alkanolamines	IonPac AS20	4.0 mm ×250 mm, 7.5 μm	Gradient elution by 5.0 mM 47.0 mM potassium hydroxide solutions 5.0 mM (-7.0 min) → 5.0 mM (0.0 min) → 5.0 mM (6.0 min) → 47.0 mM (25.0 min) → 47.0 mM (30.0 min)	1.5 mL/min	25 μL	35℃	_	
Quaternary alkanolamines	IonPac ASI 1-HC	4.0 mm×250 mm, 9 μm	Gradient elution by 1.0 mM-70 mM potassium hydroxide solutions 70.0 mM (-10.5 min) → 70.0 mM (-10.1 min) → 15 mM (-10.0 min) → 15 mM (0.0 min) → 22 mM (10.0 min) → 22 mM (14.0 min) → 42 mM (17.5 min) → 70 mM (20.0 min) → 70 mM (25.0 min) → 1 mM (25.1 min)	1.2 mL/min	25 μL	40℃	_	
			Gradient elution by 15 mM-50 mM potassium hydroxide solutions 15 mM (0.0 min) → 15 mM (18.0 min) → 50 mM (18.0 min) → 50 mM (20.0 min)			35℃	_	

Table 1 Example ion exchange column and measurement conditions in ion chromatography for acid-soluble sulfur



Figure 1 Chromatograms of sulfate ion (Part 1) Column and measurement conditions: "A" in the Note column in Table 1.



Figure 2 Chromatograms of sulfate ion (Part 2) Column and measurement conditions: "B" in the Note column in Table 1.



Figure 3 Chromatograms of sulfate ion (Part 3) Column and measurement conditions: "C" in the Note column in Table 1.



Figure 4 Chromatograms of sulfate ion (Part 4) Column and measurement conditions: "D" in the Note column in Table 1.